

# Harnessing microbes: a new approach to carbon sequestration in cocoa agroforestry

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**Keywords:** carbon sequestration; cocoa; *Trichoderma*; *Bacillus*; soil microbiology; agroforestry.

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**Abstract.** This study evaluated the carbon sequestration potential of fungal and bacterial strains in cocoa plantation soils in Ecuador's coastal region using a randomized complete block design. Four microbial treatments were tested: *Trichoderma longibrachiatum* (T1), *Trichoderma reesei* (T2), *Bacillus subtilis* (T3) and *Bacillus licheniformis* (T4). All microorganisms showed high viability, with bacterial and fungal colony-forming units exceeding  $10^8$  and  $10^6$  CFU/ml, respectively. T. longibrachiatum and T.



*T. reesei* significantly outperformed the bacterial treatments in carbon sequestration ( $p<0.0001$ ). *T. reesei* achieved a 29% increase in carbon sequestration after the first application, while *B. subtilis* showed an 11.25% increase after four applications, though with decreasing efficacy. *B. licheniformis* maintained  $\text{NH}_4^+$  at 19.00 ppm, Zn at 5.60 ppm, Mn at 5.20 ppm, and B at 0.61 ppm, while increasing P to 66.00 ppm, K to 1.89 Meq/100ml, Ca to 19.00 Meq/100ml, Mg to 5.10 Meq/100ml, S to 50.00 ppm, Cu to 5.20 ppm, and Fe to 54.00 ppm. Future research should focus on optimizing microbial dosages and application methods to enhance carbon capture and cocoa productivity.

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## 1. Introduction

Greenhouse gas emissions from agriculture, primarily from livestock methane, are a significant contributor to climate change, accounting for approximately 18% of global emissions (Belezaca et al., 2022). Latin America, including Ecuador, is a major source of these emissions, with the agricultural sector being a key contributor (SELLCA, 2024; Paccha et al., 2023). Ecuador's agricultural practices, often characterized by deforestation and excessive chemical use, exacerbate this issue (Racines, 2018; FAO, 2022). To mitigate these emissions, carbon sequestration in soil and plant biomass offers a promising strategy (Ortiz & Batioja, 2023). Forests and agroforestry systems, including cocoa cultivation, have demonstrated significant carbon capture potential (Leiva & Ramírez, 2021; Buitrago et al., 2018; Andrade et al., 2020), with soil microorganisms playing a crucial role in this process (Ahmed et al., 2019).

This study introduces a novel approach by focusing on the carbon sequestration potential of specific fungal and bacterial strains within cocoa plantation soils in Ecuador's coastal regions. Unlike previous research that broadly examines microbial impacts, our study aims to provide a detailed assessment of how *Trichoderma* and *Bacillus* strains can enhance carbon capture in this context-specific setting. By evaluating the viability and efficacy of these microorganisms, our research seeks to contribute new insights into optimizing microbial applications for improved soil health and carbon storage. The findings are intended to bridge gaps in the existing literature by offering practical recommendations for integrating microbial inoculation into cocoa

agroecosystems, with the potential to enhance sustainability and contribute to multiple Sustainable Development Goals (SDGs), including climate action, life on land, zero hunger, economic growth, and partnerships for the goals.

The potential applications and positive societal outcomes of this research lie in demonstrating how specific microbial strains can enhance carbon sequestration in cocoa plantations. By providing actionable strategies for farmers and policymakers, the study aims to support efforts to reduce agricultural emissions and promote sustainable practices. Although the results are context-specific to Ecuador's coastal cocoa plantations, they offer a valuable model for similar agroforestry systems, paving the way for broader implementation and environmental benefits. Ultimately, this study evaluates the carbon sequestration potential of selected fungal and bacterial strains, contributing to the development of targeted strategies for improving soil health and advancing sustainability in cocoa cultivation.

## 2. Methodology

### 2.1. Description of the study area

This research was conducted at the Escuela Superior Politécnica Agropecuaria de Manabí Manuel Félix López (ESPAM MFL), a higher education institution situated in Bolívar Canton, Manabí Province, Ecuador ( $0^{\circ}49'8.87''S$ ,  $80^{\circ}10'53.03''W$ , figure 1). Located at 15 meters above sea level, the region experiences a tropical climate with average temperatures ranging from  $20.60^{\circ}C$  to  $31.11^{\circ}C$ , annual rainfall of 624 mm, and relative humidity of 82.42%.

### 2.2. Microorganism activation

A completely randomized design (CRD) was employed for this study, with four treatments and four replications per treatment, totaling 16 experimental units. Each treatment consisted of the application of a specific fungal or bacterial strain (Table 1).

#### Fungal activation

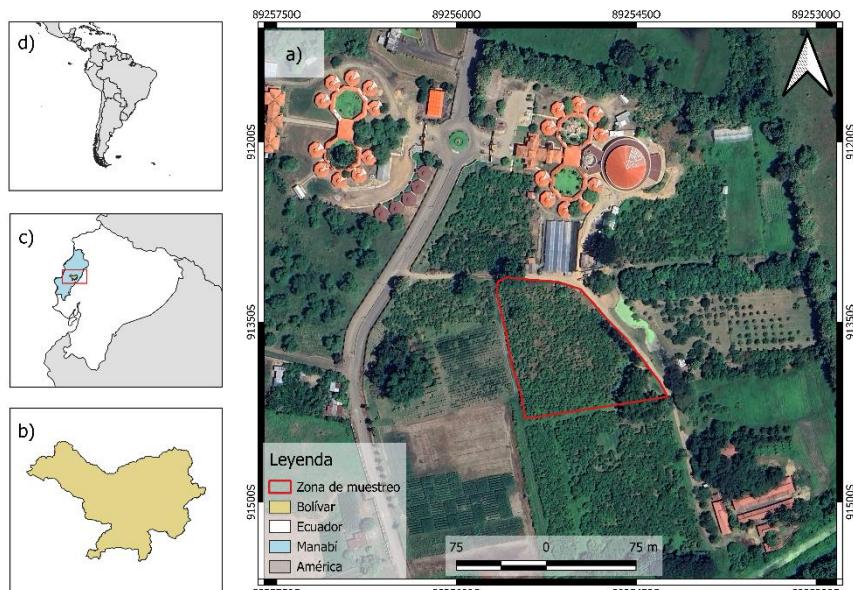
Fungal strains were cultured on Potato Dextrose Agar (PDA) in Petri dishes. A sterile inoculation loop was used to transfer fungal suspensions to the plates, which were subsequently incubated in the dark at room temperature for 24 hours. Following incubation, fungal colony growth and development were assessed.

#### Bacterial activation

Bacteria were initially cultured in a liquid peptone water medium before being transferred to nutrient agar plates. These plates were incubated at 37°C for 24 hours under aerobic conditions. Bacterial colony growth and development were monitored.

#### Microorganism reproduction

To increase microorganism populations, serial dilutions were prepared according to the protocol outlined by the Ministry of Agriculture and Livestock of Ecuador (2014) and Mero (2019). Briefly, activated microorganisms were suspended in sterile water, centrifuged, and resuspended in peptone water. Serial dilutions ranging from  $10^6$  to  $10^8$  CFU/mL were prepared.



**Figure 1.** a) Geographic location of the sampling area, b) Bolívar, c) Ecuador, d) South America.

#### *2.3. Soil analysis*

To assess soil quality, samples were collected from ESPAM MFL cocoa plantations before and seven days after experimental treatments. These samples were submitted to the Instituto Nacional de Investigaciones Agropecuarias of

Ecuador (INIA) for analysis of pH, organic matter, NH<sub>4</sub>, P, K, Ca, Mg, S, Zn, Cu, Fe, Mn, and B. Standard laboratory methods were employed, including potentiometry, Walkley-Black, atomic absorption, modified Olsen, micro Kjeldahl, and Yuan procedures (Table 2).

**Table 1.** Applied treatments.

| Treatment   | Code | Concentration              |
|---|------|----------------------------|
| <i>Trichoderma longibrachiatum</i>                  | T1   | 1 x 10 <sup>8</sup> ufc/ml |
| <i>Trichoderma reesei</i>                           | T2   | 1 x 10 <sup>6</sup> ufc/ml |
| <i>Bacillus subtilis</i>                            | T3   | 1 x 10 <sup>8</sup> ufc/ml |
| <i>Bacillus licheniformis</i>                       | T4   | 1 x 10 <sup>7</sup> ufc/ml |
| Dilution concentration : 1 x 10 <sup>8</sup> ufc/ml |      |                            |

**Table 2.** Parameters analyzed in the study and their classification criteria.

| Parameter       | Class criteria |             |             | Unit      |
|-----------------|----------------|-------------|-------------|-----------|
|                 | High           | Medium      | Low         |           |
| NH <sub>4</sub> | > 30,1         | 12,1 - 30,0 | < 12,0      |           |
| P               | > 15,0         | 8,0 - 14,0  | < 1,0 - 7,0 | ppm       |
| K               | > 0,4          | 0,2 - 0,4   | < 0,2       |           |
| Ca              | > 4,1          | 2,0 - 4,0   | < 2,0       | Meq/100ml |
| Mg              | > 2,1          | 0,8 - 2,0   | < 0,8       |           |
| S               | > 16,0         | 5,0 - 15,0  | < 5,0       |           |
| Zn              | > 7,1          | 3,1 - 7,0   | 3,0         |           |
| Cu              | > 4,1          | 1,1 - 4,0   | 1,0         |           |
| Fe              | > 41,0         | 21,0 - 40,0 | 20,0        | ppm       |
| Mn              | > 15,1         | 5,1 - 15,0  | 5,0         |           |
| B               | > 0,6          | 0,2 - 0,6   | < 0,2       |           |
| Organic matter  | > 6,1          | 6,0 - 3,1   | < 3,0       | %         |

#### 2.4. Carbon uptake assessment

Carbon uptake was determined by measuring soil respiration rates (CO<sub>2</sub> release) before and after treatment application. Laboratory analyses, including titration and CO<sub>2</sub> estimation, were conducted to assess soil biological activity, carbon storage, and the impact of treatments on carbon sequestration.

### 3. Results and Discussion

#### Microorganism viability

Both fungal (*Trichoderma longibrachiatum* and *Trichoderma reesei*) and bacterial (*Bacillus subtilis* and *Bacillus licheniformis*) strains exhibited high viability, consistently exceeding  $10^6$  and  $10^8$  CFU/mL, respectively. These findings align with previous research by Bolaños et al. (2014); Bampidis et al. (2023); Hernández et al. (2019); González et al. (2023).

#### Nutrient availability

*Trichoderma* and *Bacillus* application significantly enhanced soil nutrient availability. Initially low levels of NH<sub>4</sub>, Zn, Fe, and Mn increased following treatments, with NH<sub>4</sub> and Zn reaching medium levels and Fe attaining high levels. These results corroborate findings by Abdelmoaty et al. (2022); Andrade et al. (2023) regarding Trichoderma's role in organic matter decomposition and nutrient release. *Bacillus* demonstrated a more limited effect on Mn, increasing it only to medium levels. Mg, S, and Cu levels increased to high levels with both treatments, while B exhibited variable responses. P, K, Ca, and organic matter remained consistently high. Araújo et al. (2022) support the hypothesis of increased nutrient use efficiency following microbial inoculation. Soil pH was neutralized from 6.7 to 7.3 in all treatments (table 3).

**Table 3.** Results of soil parameter analysis before and after each treatment.

| Parameter       | Pre application | T1<br>( <i>T. longibrachiatum</i> ) | T2<br>( <i>T. reesei</i> ) | T3<br>( <i>B. subtilis</i> ) | T4<br>( <i>B. licheniformis</i> ) |
|-----------------|-----------------|-------------------------------------|----------------------------|------------------------------|-----------------------------------|
| NH <sub>4</sub> | 8,00 L          | 29,00 M                             | 22,00 M                    | 24,00 M                      | 19,00 M                           |
| P               | 66,00 H         | 57,00 H                             | 57,00 H                    | 56,00 H                      | 66,00 H                           |
| K               | 0,98 H          | 1,79 H                              | 1,87 H                     | 2,00 H                       | 1,89 H                            |
| Ca              | 11,00 H         | 18,00 H                             | 18,00 H                    | 19,00 H                      | 19,00 H                           |
| Mg              | 1,00 M          | 4,10 H                              | 5,40 H                     | 4,70 H                       | 5,10 H                            |
| S               | 15,00 M         | 55,00 H                             | 60,00 H                    | 55,00 H                      | 50,00 H                           |
| Zn              | 1,00 L          | 5,80 M                              | 6,10 M                     | 6,00 M                       | 5,60 M                            |
| Cu              | 2,60 M          | 4,80 H                              | 4,70 H                     | 4,40 H                       | 5,20 H                            |
| Fe              | 13,00 L         | 54,00 H                             | 56,00 H                    | 46,00 H                      | 54,00 H                           |
| Mn              | 3,20 L          | 4,80 L                              | 4,90 L                     | 5,80 M                       | 5,20 M                            |
| B               | 0,55 M          | 0,98 H                              | 0,57 M                     | 0,30 L                       | 0,61 M                            |
| Organic matter  | 7,00 H          | 8,00 H                              | 7,90 H                     | 8,00 H                       | 7,80 H                            |
| pH              | 6,7 H           | 7,3 N                               | 7,3 N                      | 7,3 N                        | 7,3 N                             |

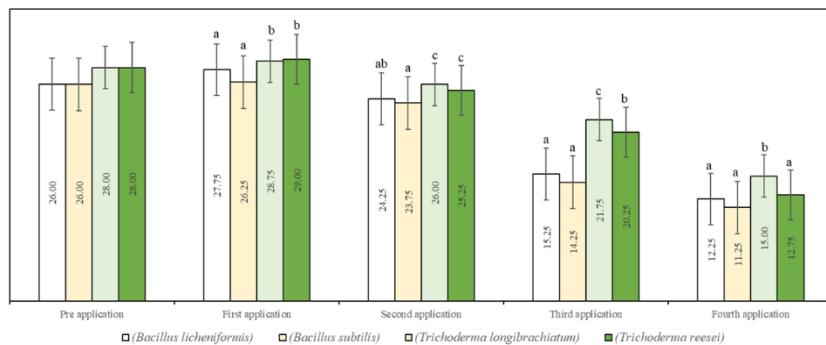
\*H=High; M=Medium; L=Low (criteria in Table 2).

All microbial treatments significantly enhanced soil carbon sequestration ( $p<0.05$ ) compared to the control. *Trichoderma reesei* initially exhibited the highest carbon capture rate (29.00 mg\*kg-s), followed closely by *Trichoderma*

*longibrachiatum*. *Bacillus subtilis* and *Bacillus licheniformis* showed lower carbon sequestration rates. These findings align with previous research highlighting the potential of *Trichoderma* species for carbon sequestration (Obando & Vélez, 2023; Conrado et al., 2019; González et al., 2023).

While *Trichoderma longibrachiatum* maintained a consistent carbon capture throughout the experimental period, *Trichoderma reesei* exhibited a slight decline after the initial peak. Both *Bacillus* species showed limited carbon sequestration capacity (figure 2). Although both *Trichoderma* and *Bacillus* species are beneficial soil microorganisms, our results indicate that *Trichoderma*, particularly *Trichoderma longibrachiatum*, is more effective in promoting carbon sequestration under the studied conditions. While Mohammed et al. (2017) reported the positive impact of these microorganisms on plant growth and disease control, our study focused specifically on their carbon sequestration potential.

The results obtained suggest that the use of *Trichoderma longibrachiatum* as a biofertilizer could be a promising strategy to increase carbon storage in agricultural soils, thus contributing to mitigating the effects of climate change. Further studies are recommended to evaluate the long-term impact of this microorganism on soil carbon dynamics and its interaction with other soil factors.



**Figure 2.** Carbon sequestration rates ( $\text{mg CO}_2\text{-C kg}^{-1}$  soil) among treatments over four weeks. Error bars represent standard error. Means with the same letter are not significantly different ( $p > 0.05$ ) according to Tukey's HSD test.

*Trichoderma longibrachiatum* and *Trichoderma reesei* demonstrated superior carbon sequestration capabilities compared to *Bacillus subtilis* and *Bacillus licheniformis* throughout the four-week study. However, a general decline in carbon capture was observed across all treatments after the initial week. This reduction is likely attributed to factors such as plant and microbial carbon uptake, as well as ongoing soil organic matter decomposition (Pérez et al., 2021; Zamora et al., 2020).

While *Trichoderma* treatments showed promise in enhancing carbon sequestration, the results indicate the need for further research to optimize application rates and assess long-term effects. Additionally, exploring alternative strategies to sustain carbon capture in cocoa plantation soils is warranted. The observed decrease in carbon sequestration compared to initial levels and findings from previous studies (Ortiz, 2019; Barrezueta, 2019) suggests that cocoa agroecosystems may require specific carbon management practices to maintain soil fertility and productivity.

#### 4. Conclusions

A comprehensive analysis of the research data indicates that *Trichoderma* and *Bacillus* strains exhibited high viability, with bacterial and fungal colony-forming units exceeding  $10^8$  and  $10^6$  CFU/ml, respectively, suggesting their potential for successful establishment in field conditions. Although microbial inoculation led to initial improvements in soil properties. Such as *Bacillus licheniformis* maintaining  $\text{NH}_4^+$  at 19.00 ppm, Zn at 5.60 ppm, Mn at 5.20 ppm and B at 0.61 ppm, while increasing P to 66.00 ppm, K to 1.89 Meq/100ml, Ca to 19.00 Meq/100ml, Mg to 5.10 Meq/100ml, S to 50.00 ppm, Cu to 5.20 ppm and Fe to 54.00 pp, the overall impact on carbon sequestration was limited. Specifically, *Trichoderma reesei* achieved a 29% increase in carbon sequestration after the first application, while *Bacillus subtilis* showed an 11.25% increase after four applications, though with diminishing returns. These findings suggest that while microbial inoculation can positively influence soil health, a more holistic approach is necessary to achieve sustained carbon storage within complex cocoa agroecosystems. To optimize microbial applications and maximize their benefits, future research should focus on integrated management strategies that consider the specific conditions of cocoa plantations and the intricate interactions within the soil ecosystem.

## References

- Abdelmoaty, S., Khandaker, M., Mahmud, K., Majrashi, A., Alenazi, M. y Badaluddin, N. (2022). Influence of Trichoderma harzianum and Bacillus thuringiensis with reducing rates of NPK on growth, physiology, and fruit quality of Citrus aurantifolia. *Brazilian Journal of Biology*, 82, 1-14. <https://doi.org/10.1590/1519-6984.261032>
- Ahmed, AAQ, Odelade, KA, Babalola, OO (2019). Inoculantes microbianos para mejorar el secuestro de carbono en agroecosistemas para mitigar el cambio climático. En: Leal Filho, W. (eds) Handbook of Climate Change Resilience. Springer, Cham. [https://doi.org/10.1007/978-3-319-71025-9\\_119-1](https://doi.org/10.1007/978-3-319-71025-9_119-1)
- Agencia de Regulación y Control Fito y Zoosanitario (Agrocalidad, 2018). Muestreo para análisis de suelo. <https://www.agrocalidad.gob.ec/wp-content/uploads/2020/05/agua8.pdf>
- Andrade, P., Rivera, M., Landero, N., Silva, H., Martínez, S. y Romero, O. (2023). Ecological and biological benefits of the cosmopolitan fungus Trichoderma spp. in agriculture: A perspective in the Mexican countryside. *Revista Argentina de Microbiología*, 55, 366-377. <https://www.elsevier.es/es-revista-revista-argentina-microbiologia-372-pdf-S0325754123000603>
- Araújo, S., Figueiredo, A., Salgado, G., Días, D., Carvalho, T. y Barata, G. (2022) Co-Inoculation of Trichoderma asperellum with Bacillus subtilis to Promote Growth and Nutrient Absorption in Marandu Grass. *Applied and Environmental Soil Science*, 2022, 1-13. <https://doi.org/10.1155/2022/3228594>
- Bampidis, V., Azimonti, G., De Lourdes Bastos, M., Christensen, H., Dusemund, B., Durjava, M. F., Kouba, M., López-Alonso, M., Puente, S. L., Marcon, F., Mayo, B., Pechová, A., Petkova, M., Ramos, F., Sanz, Y., Villa, R. E., Woutersen, R., Dierick, N., Saarela, M., y Anguita, M. (2023b). Safety and efficacy of a feed additive consisting of endo-1,4-beta-xylanase produced by Trichoderma reesei ATCC PTA-5588, protease produced by Bacillus subtilis CBS 148232, and alpha-amylase produced by Bacillus licheniformis ATCC SD-6525 (Axtra® XAP 104 TPT) for chickens for fattening, laying hens and minor poultry species (Genencor international B.V.). *EFSÁ Journal*, 21(2). <https://doi.org/10.2903/j.efsa.2023.7816>
- Barrezueta, S. (2019). Propiedades de algunos suelos cultivados con cacao en la provincia El Oro, Ecuador. *Revista Ciencia UAT (Universidad Autónoma de Tamaulipas)*, 14(1), 155-166. <https://doi.org/10.29059/cienciauat.v14i1.1210>
- Bazán, T. (2017). Manual de procedimientos de los análisis de suelos y agua con fines de riego. Universidad Nacional Agraria la Molina, Instituto Nacional de Innovación Agraria. Lima Perú. 92 p. [http://repositorio.inia.gob.pe/bitstream/inia/504/1/BazanManual\\_de\\_procedimientos\\_de\\_los.pdf](http://repositorio.inia.gob.pe/bitstream/inia/504/1/BazanManual_de_procedimientos_de_los.pdf).

- Belezaca, C., Morales, C., Solano, E. y Díaz, N. (2022). Emisiones de CO<sub>2</sub> y contenidos de carbono de la biomasa microbiana del suelo en el “Bosque Protector Murocomba”, occidente de los Andes Ecuatorianos. *Revista de Investigación, Talentos*, 9(1), 18-32. <https://doi.org/10.33789/talentos.9.1.158>
- Bolaños, B., González, H., Zavaleta, E., Sánchez, P., Mora, G., Nava, C., Real, J., y Rubio, R. (2014). Colonización de Trichoderma y Bacillus en Plántulas de Agave tequilana Weber, var. Azul y el Efecto Sobre la Fisiología de la Planta y Densidad de Fusarium. *Revista mexicana de fitopatología*, 32(1), 62-74.  
[https://www.scielo.org.mx/scielo.php?pid=S0185-33092014000100006&script=sci\\_arttext](https://www.scielo.org.mx/scielo.php?pid=S0185-33092014000100006&script=sci_arttext)
- Burbano, H. (2018). El carbono orgánico del suelo y su papel frente al cambio climático. *Revista de Ciencias Agrícolas*, 35(1), 82-96.  
<https://doi.org/10.22267/rca.183501.85>
- Conrado, M., Mazaro, S., y Silva, J. (2019). Trichoderma: uso en la agricultura. Embrapa Soja. Brasil.
- González, Y., Ortega, J., Anducho, M y Mercado, Y. (2022). Bacillus subtilis y Trichoderma: Características generales y su aplicación en la agricultura. TIP. *Revista especializada en ciencias químico-biológicas*, 25, e520.  
<https://doi.org/10.22201/fesz.23958723e.2022.520>
- Hartmann, M., Six, J. (2023). Soil structure and microbiome functions in agroecosystems. *Nat Rev Earth Environ*, 4, 4–18. <https://doi.org/10.1038/s43017-022-00366-w>
- Hernández, D., Ferrera, R., y Alarcón, A. (2019). Trichoderma: importancia agrícola, biotecnológica, y sistemas de fermentación para producir biomasa y enzimas de interés industrial. *Chilean journal of agricultural & animal sciences*, 35(1), 98-112.  
<https://dx.doi.org/10.4067/S0719-38902019005000205>
- Leiva, E. y Ramírez, R. (2021). Carbono almacenado en cacao y suelo en sistemas agroforestales. *Brazilian Journal of Animal and Environmental Research*, 4(4), 5331-5346. DOI: 10.34188/bjaerv4n4-036
- Mero, A. (2019). *Evaluación de la incorporación de Lactobacillus brevis encapsulado en el alimento sobre los parámetros productivos, salud de pollos Cobb 500*. [Tesis de Pregrado, Escuela Superior Politécnica Agropecuaria de Manabí]. Repositorio Institucional.  
<https://repositorio.espam.edu.ec/bitstream/42000/1182/1/TIMV9.pdf>
- Ministerio de Agricultura y Ganadería. (2014). Protocolo para la reproducción de capas nativas de Trichoderma spp. en laboratorios artesanales.  
<https://www.agricultura.gob.ec/wp-content/uploads/2016/01/MANUAL-labos-para-web.pdf>
- Mohammed, A. M., Robinson, J. S., Midmore, D. J., y Verhoef, A. (2017). Carbon storage in Ghanaian cocoa ecosystems. *Carbon Balance and Management*, 11(1).  
<https://doi.org/10.1186/s13021-016-0045-x>

- Obando, P y Vélez, M. (2023). *Evaluación de la captación de carbono mediante microorganismos en plantaciones de café (*Coffea arabica*)*. [Tesis de Pregrado, Escuela Superior Agropecuaria de Manabí]. <http://repositorio.espm.edu.ec/handle/42000/2135>
- Organización de las Naciones Unidas para la Alimentación y la Agricultura [FAO]. (2022). Condiciones Climáticas y la actividad humana impactan en la degradación de la tierra, comprometiendo la seguridad alimentaria. <https://www.fao.org/ecuador/noticias/detail-events/ru/c/1141396/>
- Ortiz, J. (2019). *Cuantificación y análisis del almacenamiento de carbono en suelos gestionados bajo modelos de producción agrícola aplicados al cultivo de cacao (*Theobroma cacao L.*) en ecosistema tropical*. [Tesis de Postgrado, Universidad de Manizales]. Repositorio Institucional. <https://ridum.umanizales.edu.co/xmlui/handle/20.500.12746/3622>
- Ortiz, J. y Batioja, D. (2023). *Evaluación de beneficios del sistema agroforestal Cacao (*Theobroma cacao L.*) y tangare (*Carapa Guianensis Aubl*) en Tumaco – Nariño*. [Tesis de grado, Universidad de Nariño]. <https://sired.udesar.edu.co/9862/1/TRABAJO%20FINAL%20JENIFER%20ORTIZ%20DAJOME%20Y%20DIOGENES%20BERNARDO%20BATIOJA.pdf>
- Paccha, J., Alvarado, V., Heidinger, H. y Ramos, L. (2024). Medición de gases de efecto invernadero en suelos agrícolas y ganaderos mediante cámaras estáticas cerradas en el sector Zalapa, ciudad de Loja. *Bosques Latitud Cero*, 14(1), 137–149. <https://doi.org/10.54753/blc.v14i1.2129>
- Pastor, J., Rivas, W., Martínez, A., Márquez, E., y Campos, Y. (2015). Carbono orgánico del suelo en un gradiente altitudinal en la Península de Paraguaná, Venezuela. *Multiciencias*, 15(3), 271-280. <https://www.redalyc.org/articulo.oa?id=90444727005>
- Pérez, H., Rodríguez, I., y García, R. (2021). Secuestro de carbono por el suelo y sus fracciones en agroecosistemas tropicales de la región costa ecuatoriana. *Universidad Y Sociedad*, 13(2), 141-149. <https://rus.ucf.edu.cu/index.php/rus/article/view/1951>
- Racines, A. (2018). *Análisis de reducción de emisiones de gases de efecto invernadero mediante descomposición aeróbica de residuos industriales en mezcla con residuos pecuarios*. [Tesis de posgrado, Universidad Andina Simón Bolívar]. <https://repositorio.uasb.edu.ec/bitstream/10644/6058/1/T2552-MCCNA-Racines-Analisis.pdf>
- Sistema Económico Latinoamericano y del Caribe (SELLA). (2024). *América Latina y el Caribe aporta el 11% del total global de emisiones de carbono*. Autor.
- Torres, W. (2016). *Efecto del uso de melaza y microorganismos eficientes sobre la tasa de descomposición de la paja de trigo (*triticum ssp*) en el barrio de Nicrapampa, distrito de Independencia, Huaraz, 2015*. [Tesis de Pregrado, Universidad Nacional Santiago Antúnez de Mayolo]. Repositorio Institucional. <http://repositorio.unasam.edu.pe/handle/UNASAM/1453>

- Zabala, J., y Vega, L. (2021). Captura y almacenamiento de carbono en distintas edades del cultivo de cacao bajo sistemas agroforestales de Tingo María (1 ed.). Perú: Biblioteca Nacional del Perú. <https://www.unheval.edu.pe/portal/wp-content/uploads/2021/10/Zavala-Vega.-2021.pdf>
- Zamora, P., Mendoza, Mayra, S., Jarquín, D., Quevedo, A., y Navarro, A. (2018). El manejo del suelo en la conservación de carbono orgánico. *Revista mexicana de ciencias agrícolas*, 9(8), 1787-1799. <https://doi.org/10.29312/remexca.v9i8.1723>

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